

# Supporting Antimicrobial Stewardship from the Laboratory

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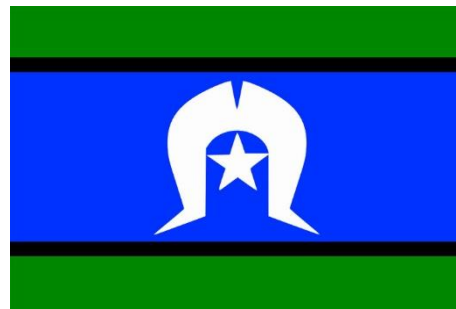
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# Acknowledgement of Country

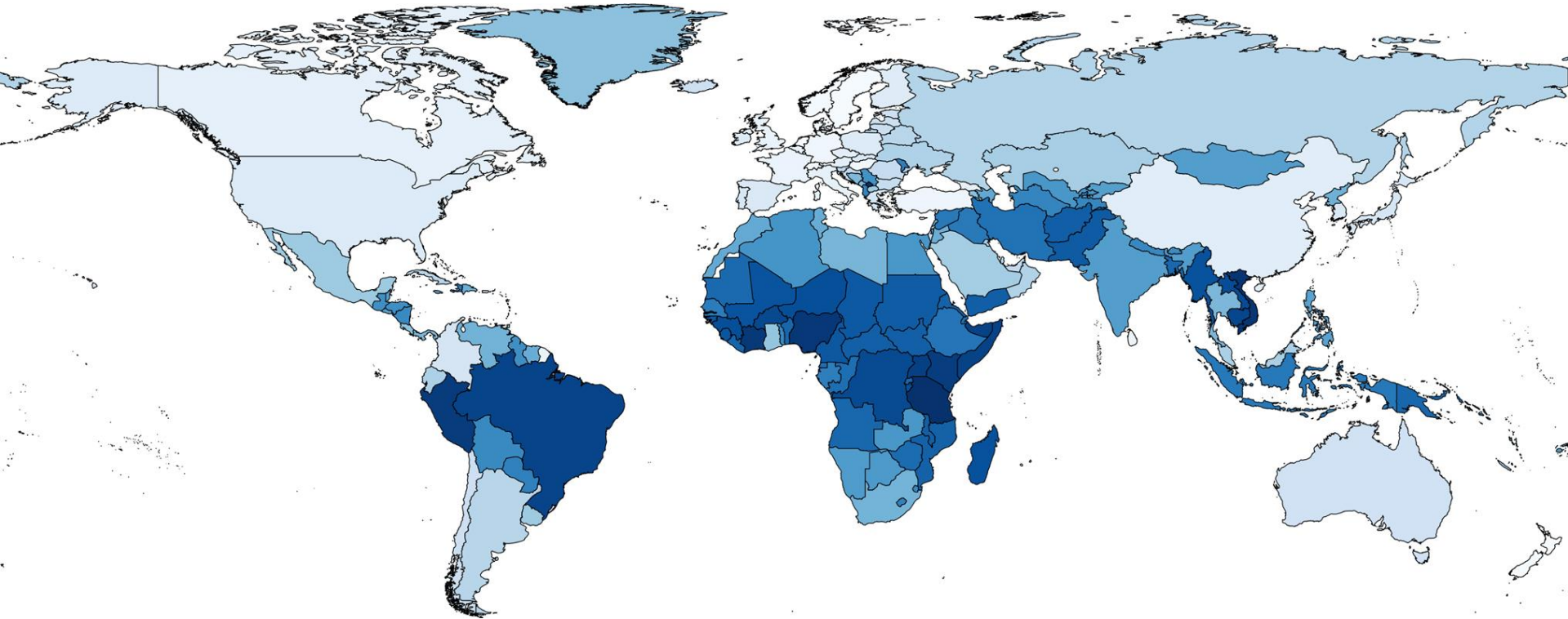
NSW Health Pathology acknowledges the traditional custodians of the lands on which we work and pay our respects to ancestors and Elders, past and present.

We are committed to honouring Australian Aboriginal and Torres Strait Islander peoples' unique cultural and spiritual relationships to the land, waters and seas and their rich contribution to society.



# Data & information

Global monitoring of antimicrobial resistance based on metagenomics analyses of urban sewage



NATURE COMMUNICATIONS | (2019)10:724 | <https://doi.org/10.1038/s41467-019-08853-3> | [www.nature.com/naturecommunications](http://www.nature.com/naturecommunications)



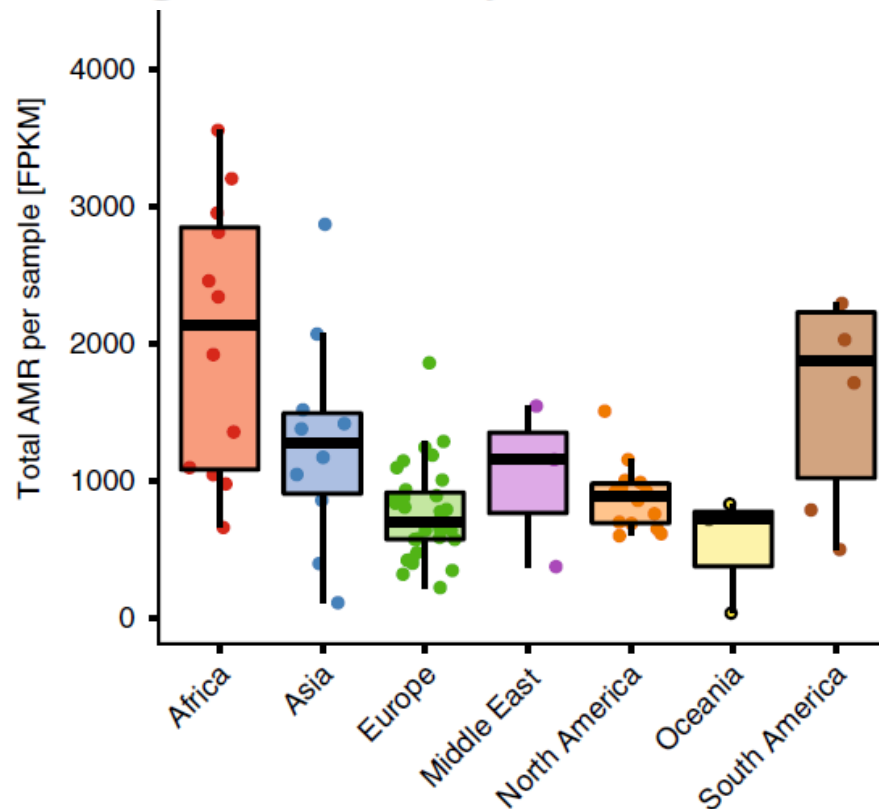
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# Data & information

Global monitoring of antimicrobial resistance based on metagenomics analyses of urban sewage



# Nature of the data

- Molecular
  - CARAlert, *mec* gene detection
- Minimum inhibitory concentrations
  - Disk zones
- Epidemiological cut-offs (ECOFFs)
- Breakpoints



# DATA

## Minimum inhibitory concentrations

- Standardised inoculum
  - Broth: “McFarland 0.5”  $\approx 1.5 \times 10^8$  CFU /ml
- 2 fold dilutions of antimicrobial in mg/L
  - Approximately log linear distribution of isolates

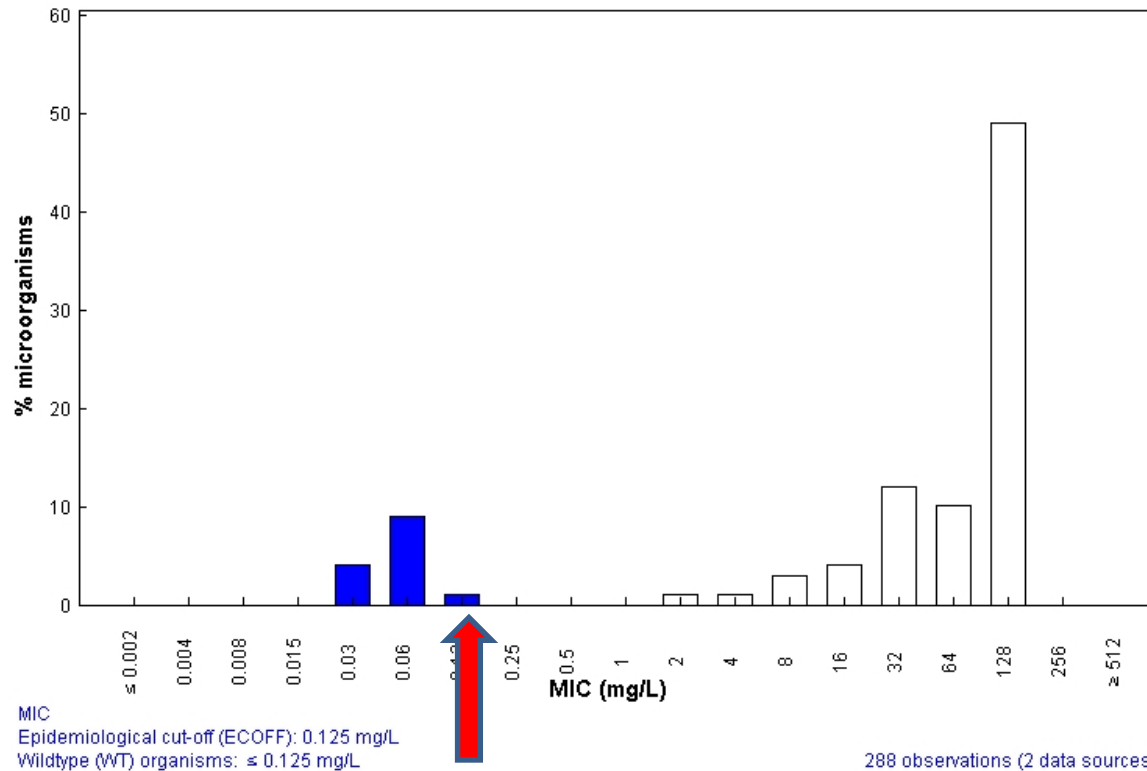


# DATA

## MIC & ECOFF

### Benzylpenicillin / *Staphylococcus aureus* MSSA International MIC Distribution - Reference Database 2019-06-19

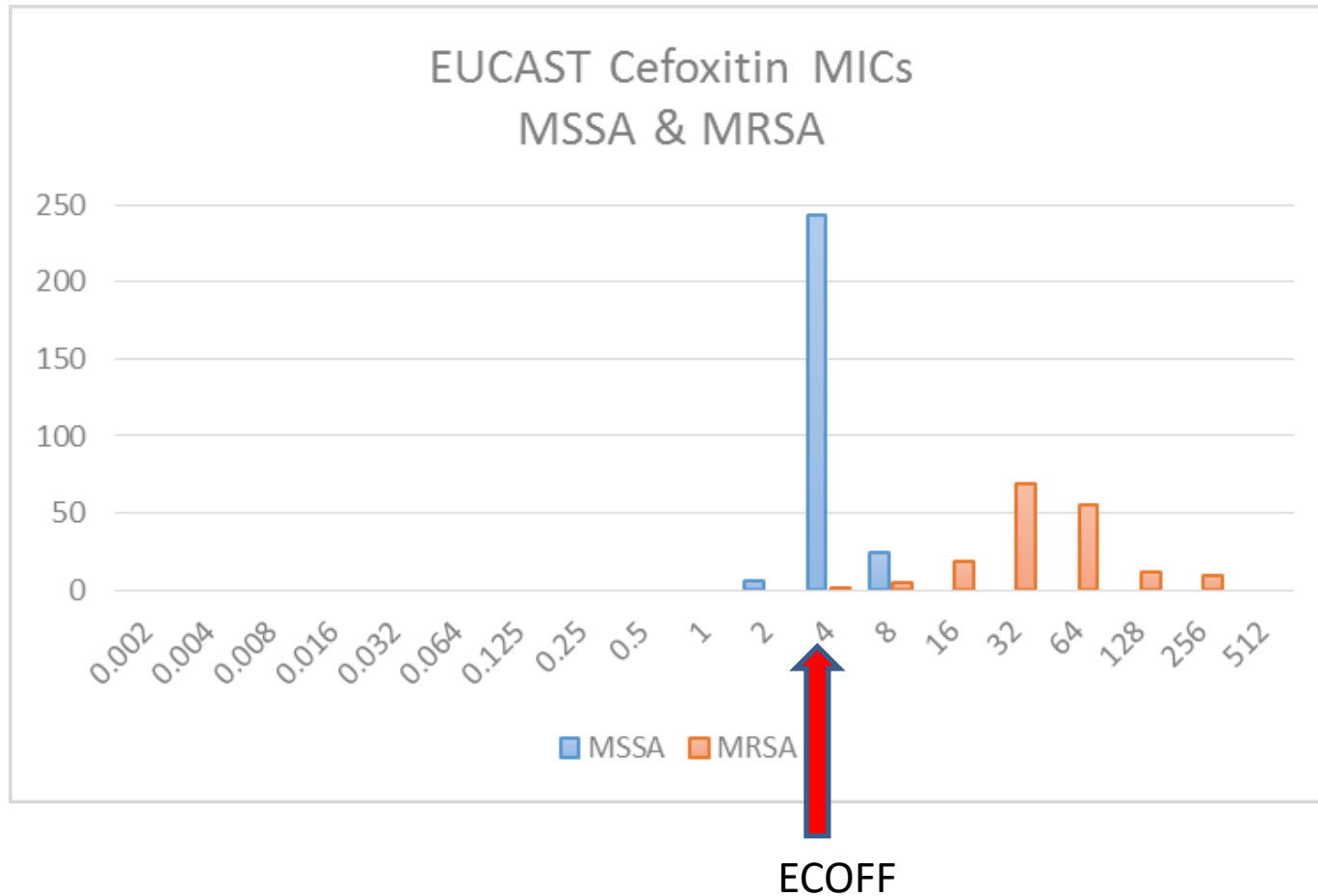
MIC distributions include collated data from multiple sources, geographical areas and time periods and can never be used to infer rates of resistance



ECOFF & Breakpoint

# DATA

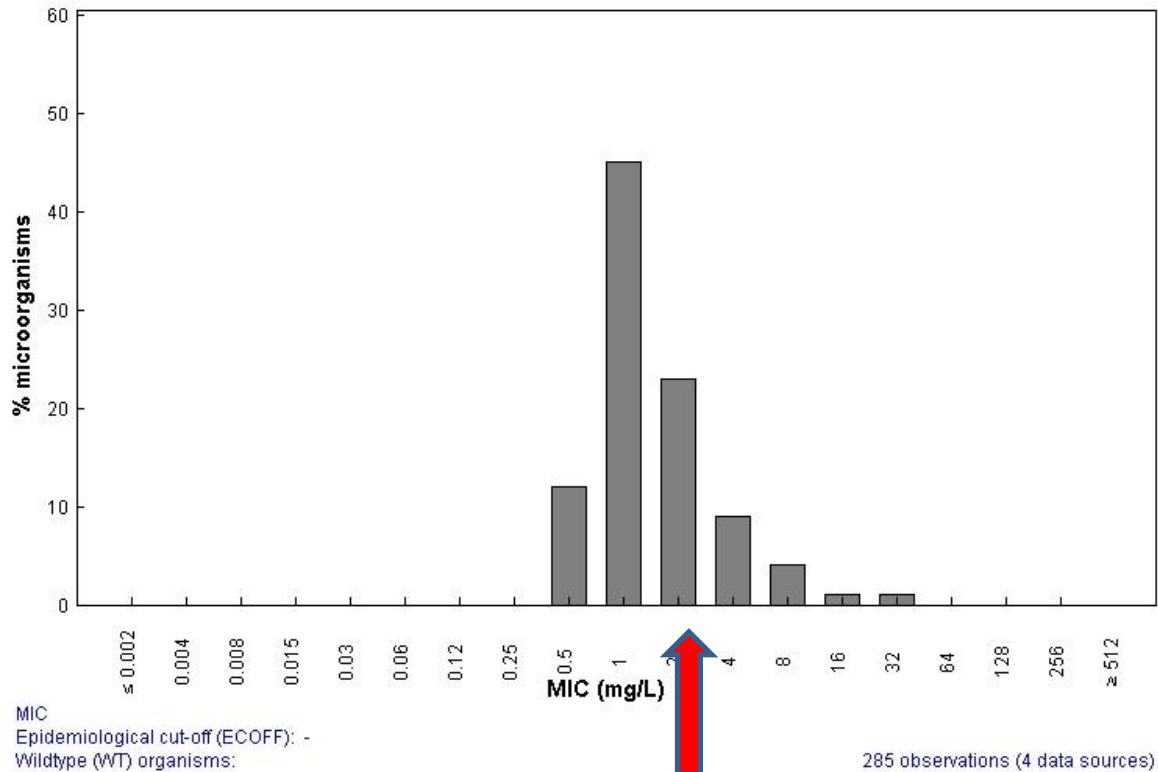
## MICs, ECOFFs & breakpoints





**Cefazolin / Escherichia coli**  
**International MIC Distribution - Reference Database 2019-06-19**

MIC distributions include collated data from multiple sources, geographical areas and time periods and can never be used to infer rates of resistance

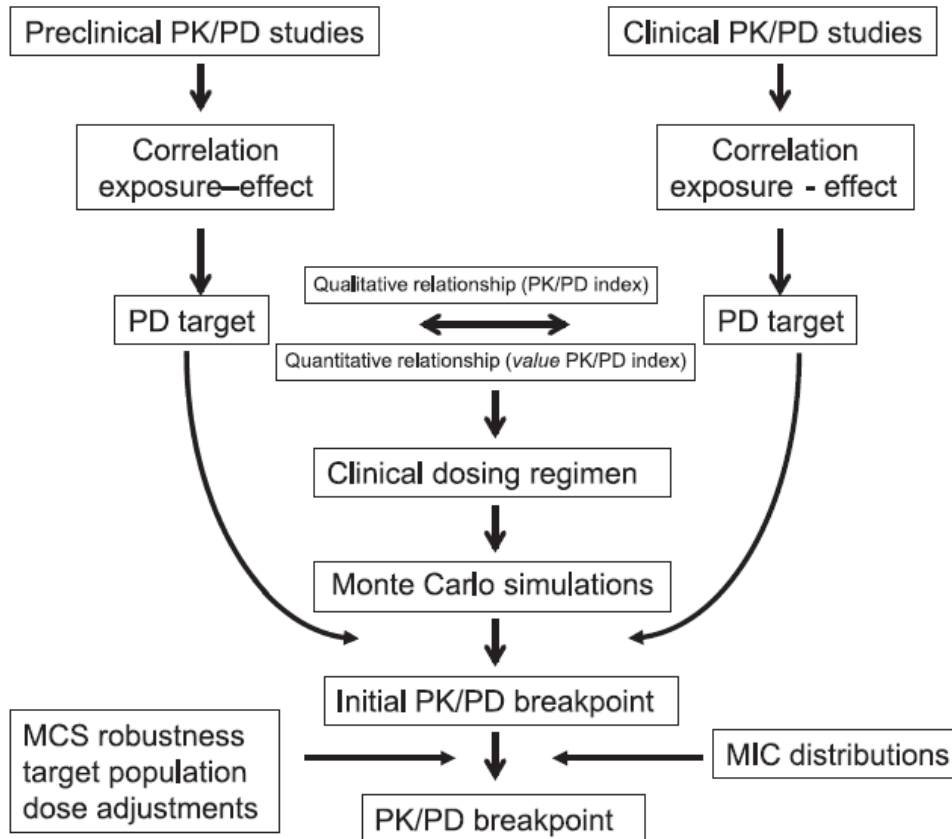


MIC  
Epidemiological cut-off (ECOFF): -  
Wildtype (WT) organisms:

**Breakpoint**

# DATA

## PK / PD & breakpoints



Mouton et al. Clin Microbiol Infect 2012; 18: E37–E45

# DATA

## Breakpoint: S or R (or I ?)

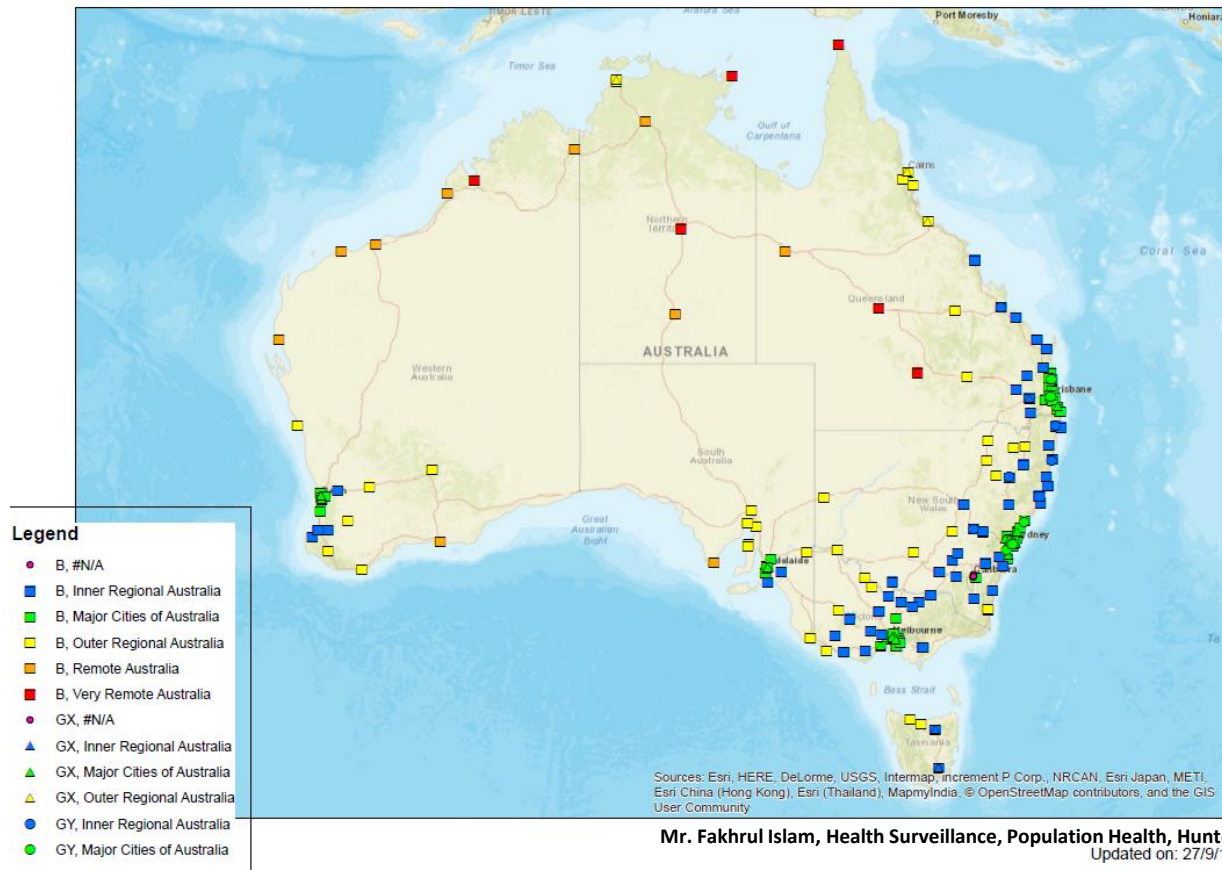
- Breakpoint determines if an isolate is classified as susceptible, resistant or intermediate
  - *unless trumped by a molecular marker*
- Three different standards for breakpoints
  - EUCAST, CLSI, CDS
  - Annual updates
- ISO 20776-1: 2006 (now 2019)
- Australian practice has been to report “intermediate” as “resistant”, but that’s about to change



# DATA

## Source of the data

Locations of NATA Accredited Microbiology Laboratories - Australia  
By Laboratory Category and Remoteness



# Turning data into information

- Antibigrams
  - Selective “cascade” reporting
- Cumulative antibigrams



# INFORMATION: ANTIBIOGRAMS

## Cascade reporting

\* Page down for extra sensitivities \*

Escherichia coli >10<sup>8</sup>/L

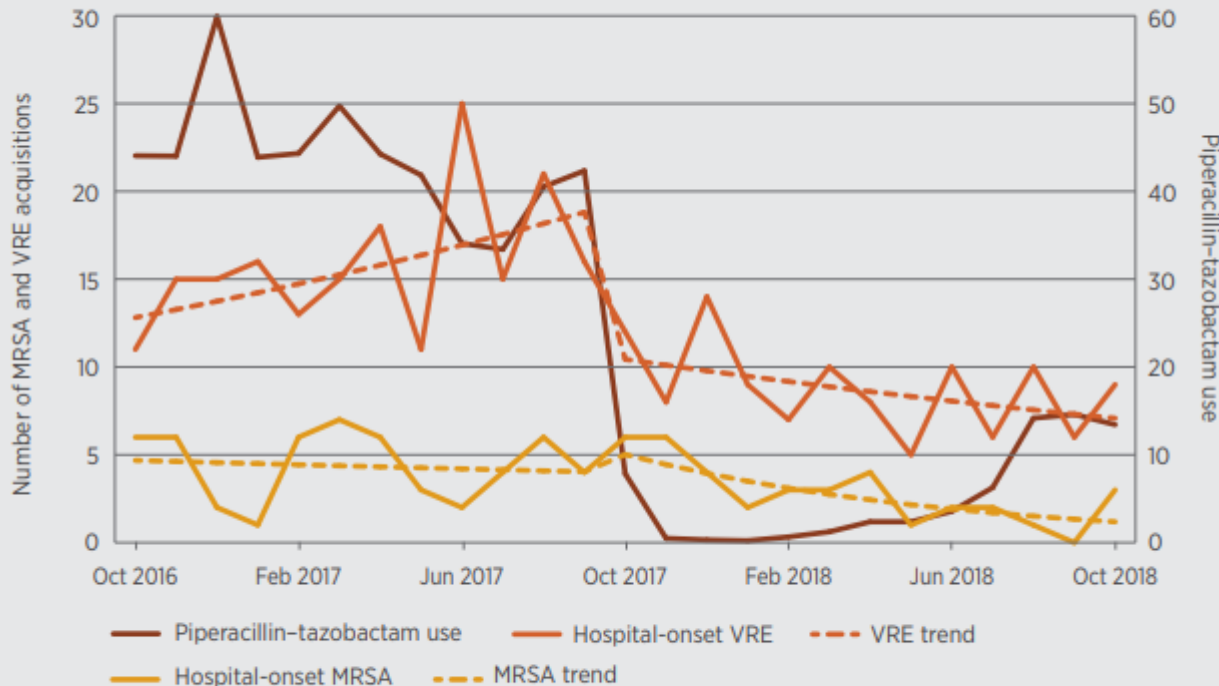
AMP	AMC	AMU	KZ	KZU	TMP	F	CRO	CAZ	TZP	GM	TOB	AK	CIP	NRX	MEP
R	S		S		R	S	S	S		S			S	S	



# INFORMATION: ANTIBIOGRAMS

## Cascade reporting

**Figure A:** Methicillin-resistant *Staphylococcus aureus* (MRSA) and vancomycin-resistant enterococci (VRE) acquisitions with changes to piperacillin-tazobactam use (DDD/1,000 OBD)



DDD/1,000 OBD = defined daily doses per 1,000 occupied bed days

Source: Ferguson JK, Kozierowski K, Munnoch SA, Oldmeadow C, Chiu S. Reduction in MRSA and VRE colonisation and infection associated with sustained reduction in broad spectrum antibiotic usage at a large tertiary hospital [article submitted to *Medical Journal of Australia*, January 2019].

# INFORMATION: ANTIBIOGRAMS

## Cascade reporting

- “We suggest selective and cascade reporting of antibiotics over reporting of all tested antibiotics (*weak recommendation, low-quality evidence*).”

Implementing an Antibiotic Stewardship Program: Guidelines by the Infectious Diseases Society of America and the Society for Healthcare Epidemiology of America CID 2016:62 (15 May)

*.....selective reporting of AST has never been evaluated using an experimental design.....”*

*Binda F, Fougnot S, De Monchy P on behalf of the ANTIBIO-CIBLÉ Scientific Committee, et al Impact of selective reporting of antibiotic susceptibility test results in urinary tract infections in the outpatient setting: a protocol for a pragmatic, prospective quasi-experimental trial BMJ Open 2018;8:e025810. doi: 10.1136/bmjopen-2018-025810*





# (DIS)INFORMATION

## Censorship

- Censored tests
  - “Pathology stewardship”
- Censored reports
  - *Stenotrophomonas maltophilia*
  - *Staphylococcus aureus*?



# INFORMATION

## Cumulative antibiograms

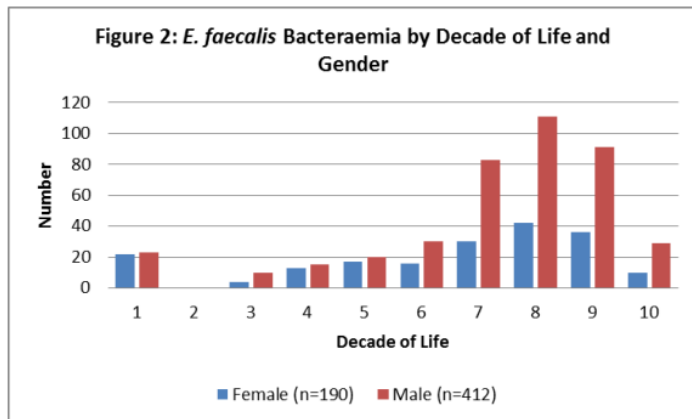
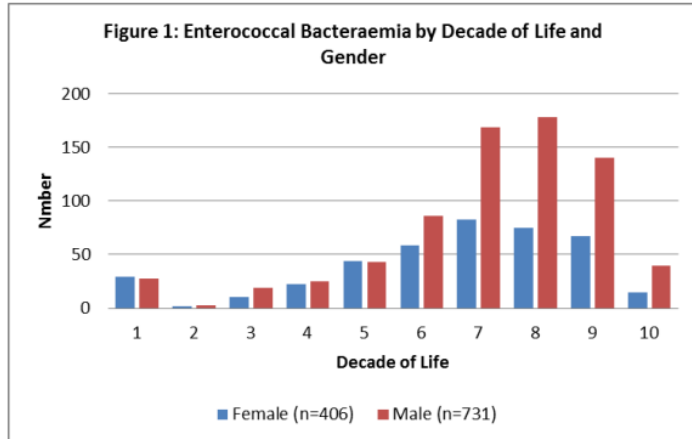
Figure A1: Example of a cumulative, hospital-level antibiogram for blood culture isolates

Organism Group	No. Organisms	%Total	Antibiotics																						
			Penicillin	Amoxicillin	Flucloxacillin	Amoxicillin-clavulanate	Piperacillin-tazobactam	Cefalotin	Cefazolin	Ceftriaxone	Ceftazidime	Cefepime	Meropenem	Gentamicin	Amikacin	Sulpha-trimethoprim	Ciprofloxacin	Fusidic Acid	Rifampicin	Gentamicin (High Level)	Erythromycin/Clarithromycin	Clindamycin	Tetracyclines	Quinupristin-dalfopristin	Vancomycin
<b>All isolates</b>	<b>2142</b>	<b>100.0</b>																							
Coagulase negative Staphylococci	637	29.7	% 11	41	40	40			R						59	79	79	98		44	64	68		100	
			n 627	621	621	621	621								629	609	623	626		626	625	628		632	
Escherichia spp	407	19.0	%	43	79	92		77	90		93	100	91	99	69	86									
			n 407	407	406	406	407	407		406	407	407	*329	407	407										
Staphylococcus aureus (ALL)	211	9.9	% 26	89	89	89			R						95	92	95	100		86	87	95		100	
			n 211	211	211	211	211							211	*185	*186	*186			211	211	211		211	
Klebsiella spp	147	6.9	%	R	94	92		84	93		95	100	99	100	86	93									
			n		147	147		147	147		147	147	147	147	*119	147	147								
viridans Streptococci	118	5.5	% 85				100	89							68		R		100	50	85	74		100	
			n 117				*15	115						25				*83	117	117	117			116	
Pseudomonas aeruginosa	100	4.7	%	R	R	89		R	R	92	94	94	96	96	R	94						R			
			n			100				100	100	100	100	*77	100										
Enterococcus spp	90	4.2	% 77				R	R	R	R					R		R	43	81	R	R	30	33	87	
			n 90														*23	*83				90	*76	90	
Spore forming GPB	57	2.7	% 17					75							92	97				76	49	78		98	
			n *54				*12								*25	*39				*21	*53	*50		*52	
Enterobacter cloacae complex	56	2.6	%	R	R	66		R	71		86	98	93	100	77	93									
			n			56		56	56	56	56	56	56	*43	56	56									

“.....it is noted that while national data is informative, local data may differ due to service provision or demographic differences in local populations. As a result, local data is the preferred reference for local AMR surveillance.” ..... “To avoid misinterpretation of antibiograms, clinical requests for access to antibiograms should be supported by concurrent consultation with an infectious diseases specialist or microbiologist

# INFORMATION

## Laboratory data *plus* epidemiology: AGAR



The Australian Group on Antimicrobial Resistance

### *E. faecium* bacteraemia: *VanA* positive among total Vancomycin resistant Enterococci

Year	Number and (%)
2013	8 of 310 (2.6%)
2014	35 of 371 (9.4%)
2015	78 of 391 (19.9%)
2016	85 of 202 (21.3%)
2017	112 of 461 (25%)

*There was no significant difference in mortality between vancomycin non-susceptible (VRE) and vancomycin susceptible E. faecium (VSE): 58/203 (29.1%) and 56/212 (26.4%) respectively (p=0.6)*

Australian Enterococcal Sepsis Outcome Program (AESOP) 2017

# Miscellaneous

- Comments on microbiology reports
- Rapid notification of organism identification and susceptibilities
- Alternative diagnoses
- *Evidence for effect ?*



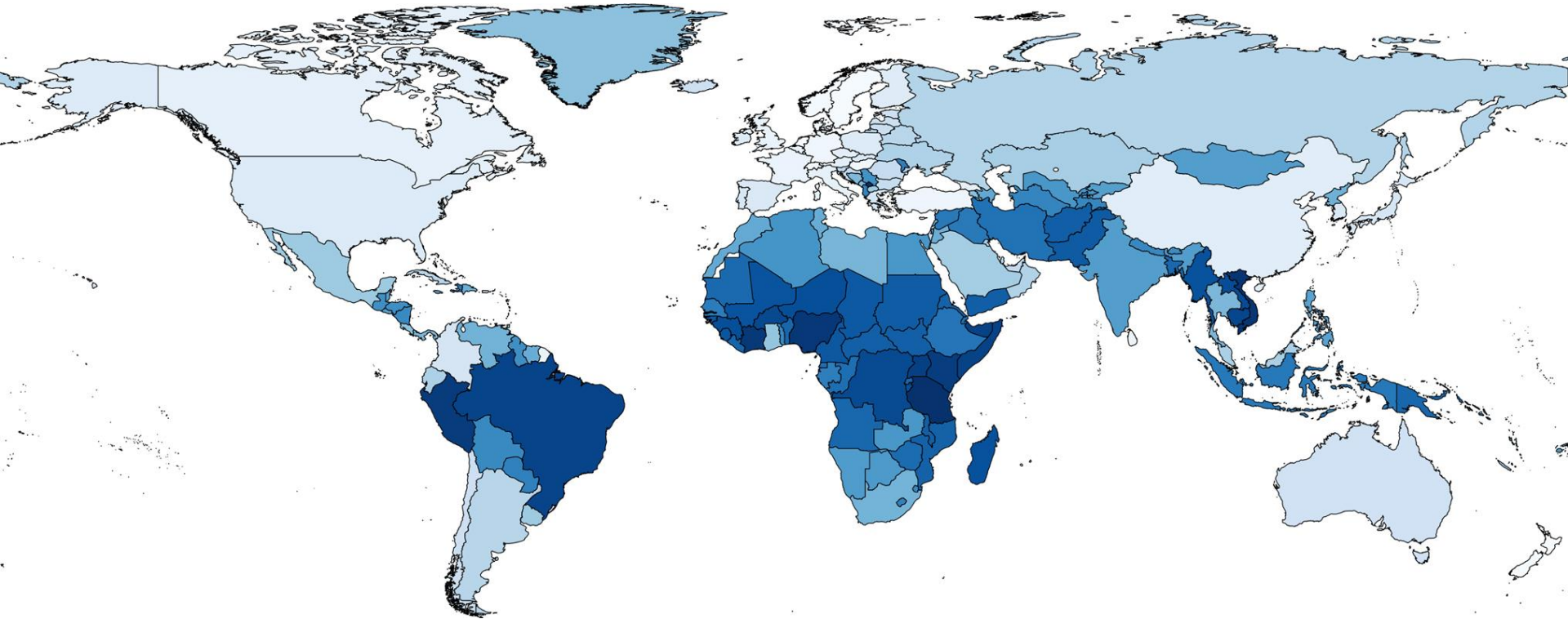
# Summary

- Data & information
- Surveillance
  - *Surveillance is data for action*
  - *Evaluation of surveillance*
  - *Speed & action*



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# Questions?



NATURE COMMUNICATIONS | (2019)10:1124 | <https://doi.org/10.1038/s41467-019-08853-3> | [www.nature.com/naturecommunications](http://www.nature.com/naturecommunications)



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